Synthesis of Copper Nanoparticles Using a Different Method: Determination of Their Antioxidant and Antimicrobial Activity

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Abstract: The aim of this study was to obtain copper oxide nanoparticles (CuO NPs) through a method of green synthesis that involves using peroxidase enzymes that are partly purified from fig leaves (Ficus carica). CuO NPs were successfully synthesized using the green synthesis method in the experiments performed. Ultraviolet–visible (UV–Vis) spectrophotometry of the characteristics of the acquired CuO NPs was performed with scanning electron microscopy (SEM) and X-ray diffraction (XRD). The optimum activation temperature for green synthesis was observed to be in 30 min, pH: 8, at 25 °C and in the concentration of 1 mM CuCl2. By using peroxidase enzymes with green synthesis, it was found that the results of the SEM and XRD measurements that acquired the CuO NPs were in the size of 50-120 nm. Afterwards, the antioxidant and antibacterial activities of these nanoparticles were measured, and it was understood from the obtained results that CuO NPs have both antioxidant and antimicrobial activities.

Keywords: Copper nanoparticles; green synthesis; peroxidase.


DOI: To be assigned.

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INTRODUCTION

Nowadays, the subject of nanotechnology encompasses many fields, including the environment, water treatment, chemical production, containers and converters of solar power, antimicrobial agents, textiles, dyes, health, and even defense [1-3]. The research indicates that nanotechnology is engaging even more with these fields as time passes.

The characteristics of a material can change significantly when that material is nano-sized. The differences in the characteristics of the substance are the result of quantum size effects. Metal nanoparticles, in particular, show very different optical, thermal, chemical, and conductivity characteristics compared to normal metal particles. Nano-sized metals and metal oxides are able to be used in many areas because of the characteristics they assume.

Green synthesis has become one of the most preferred applications in various fields, including chemistry, because of its eco-friendly approach. With the application of green synthesis to nanochemistry, another area of study had emerged that has gained increasing value: Green nano synthesis. Green nano synthesis allows for a nano material to be synthesized in a way that is friendly to both humans and to the environment. In this synthetic method, toxic chemicals that are damaging to nature are avoided and, instead, less-harmful substitutes are used. The reaction environment used in green nano synthesis can be a tissue, cell, organism, extract, enzyme, carbohydrate, lipid, or protein produced from a lipid. Three of these types of green synthesis environments are commonly used. The first of these is enzymes, which are able to catalyze a wide range of chemical reactions, resulting in their use in green nano synthesis in recent years. In addition, microorganisms (such as bacteria, yeast and mold) and plant and animal extracts are commonly used to synthesize nanoparticles in green synthesis. These three reaction environments are most common because they allow for eco-friendly, non-toxic, cost-efficient and mild conditions [6-9]. These are not, however, the only reaction environments used in green synthesis. The synthesis of silver nanoparticles (AgNPs) has been achieved by some research groups using *Nephelium lappaceum* L. and *Plectranthus amboinicus* leaf extracts. Another research group has synthesized gold nanoparticles (Au NPs) using a plant extracts in a similar way [3-4].

Copper nanoparticles, which have been a focus of interest since the end of 20th century due to their catalytic, optical, and electrical features, are widely used in the electricity sector because of their conductivity features; in chemistry, they are used as a lubricator and catalyst [10-12]. Since the copper (II) oxide nanoparticle (CuO NP) is semi-conductive, it is often used in catalysts, gas sensors, and photovoltaic cells. Due to its electronic and magnetic characteristics, CuO has been used as one of the basic components of high temperature superconductive substances, causing it to become very widely used [13]. Etefagh et al. designed a sensor founded on
nanoparticles and nanolayers of CuO. CuO NPs were produced using the sol–gel technique, and their nanolayers were prepared using spray pyrolysis [14]. Phiwdang’s research group investigated the effect of starting precursors on structural properties of CuO nanostructures synthesized via the precipitation technique [15]. Khashan et al. discovered the consequence of altering the ablation time and laser energy on produced CuO NPs using laser ablation in liquid. The antibacterial activity of these CuO NPs with or without amoxicillin on cultures of gram-negative and gram-positive bacteria was also presented [16].

Recently, fig extracts have become another preferred reaction environment for green nano synthesis. For instance, Hu (2015) achieved a synthesis of tin dioxide (SnO$_2$) using the extract obtained from fig leaves, applying the synthesized compound to the electrochemical detection of mercuric Hg(II) [3]. In another field, Singh and Bhakat studied the nano technological applications of gold and silver nanoparticles after synthesizing both with the leaves and bark of a fig tree [6].

Peroxidases are known as (EC.1.11.1.7) oxidoreductases. Usually, there is an iron porphyrin ring on their structures, and they catalyze redox reactions such as electron transfer and oxygen exits from a donor such as hydrogen peroxide (H$_2$O$_2$). The peroxidase enzyme (POX) has been purified so far in plants such as turnips, soy beans, tomatoes, potatoes, carrots, wheat, pears, apricots, bananas, dates, and strawberries [17].

In this study, we used POX that has been partly purified from the leaves of a fig plant. CuO NPs were synthesized using the purified enzymes. The characterization of the obtained CuO NPs was performed using an ultraviolet–visible (UV–VIS) spectrophotometer, X-ray diffraction (XRD), scanning electron microscopy (SEM) and Fourier transform infrared spectroscopy (FTIR). The antioxidant and antimicrobial features of the obtained CuO NPs were discovered.

**EXPERIMENTAL SECTION**

**Chemicals and Reagents**

Copper(II) chloride (CuCl$_2$), 2,2’-azino-bis (3-ethylbenzthiazol-6-sulfonic acid) (ABTS), neocuproine (2,9-dimethyl-1,10-phenanthroline), riboflavin, methionine, nitroblue tetrazolium (NBT), 1,1-diphenyl-2-picryl-hydrazyl (DPPH), 3-(2-pyridyl)-5,6-bis (4-phenyl-sulfonic acid) - 1,2,4-triazine (ferrozine), α-tocopherol, linoleic acid, gallic acid, quercetin, Folin-Ciocalteu and trichloroacetic acid (TCA) was purchased from Sigma-Aldrich GmbH (Sternhe I Germany). All other chemicals were obtained from Merck.

**Preparation of Plant Samples**

The fig plant (*Ficus carica*) used in the study was collected from Sakarya, Turkey, and it was identified with the help of taxonomists. The plant was cut into small (50 g) pieces. The pieces
were thoroughly shattered to form a homogeneous mixture in a blender, along with a 250 mL, 10 mM sodium phosphate buffer (pH:6.0). The mixture was centrifuged at 5000 g for 10 min, and the supernatant was used for enzyme purification [18].

**Partial Purification of the POX with Ammonium Sulfate Precipitation**
The upper solution of the fig plant in sodium phosphate buffer was saturated from 60% to 80% with ammonium sulfate; then, the POX was precipitated by being centrifuged at 7000 g for 30 min. The obtained precipitate was dissolved in the 5 mL, 10 mM sodium phosphate buffer (pH:6.0) and was incubated at 4°C for further analysis [18].

**POX Activity Test**
The peroxidase activity assay was measured using a substrate of 1 mM 2, 2'-azino-bis(3-ethylbenzthiazoline-sulfonic acid) diammonium salt (ABST) prepared in a 0.1 M phosphate buffer (pH:6.0). The changes in absorbance were monitored at 412 nm using a UV–Vis spectrophotometer at 1-min intervals for 3 min [18].

**Synthesis of CuO NPs**
An amount of 300 µg/mL of purified POX from the fig plant was added to a sample of copper-(II) chloride solution (CuCl₂) (2.9 mL, 10 mM) and incubated in a closed space for 4 hours. The solution became blue in color and was cloudy, indicating the presence of CuO NPs. Then, water was removed with the help of an evaporator, and the synthesized CuO NPs were dried at 70°C for 24 hours [18].

**Characterization of CuO NPs**
Synthesized CuO NPs were characterized by being scanned with a UV–VIS spectrophotometer (PG Instrument T80 UV-VIS spectrophotometer, UK) at a range of 200–1000 nm. Determination of the topography of the CuO NPs was performed using a scanning electron microscope (Sigma 300, SEM Zeiss, Germany). In addition, XRD analysis (PANalytical, Empyrean, Netherlands) was performed at DAYTAM (Eastern Anatolian High Technology Applications and Research Center) to determine the size of the CuO NPs.

**Optimization of Green Synthesis Reaction Medium**

**Optimum contact time:** To determine the optimum contact time, samples were spectrophotometrically measured between 0 and 240 min at 3-min intervals.

**Determination of optimum pH:** Synthesis of the CuO NPs was performed at pH 2–3, an acetate buffer at pH 4–6, a phosphate buffer at pH 7–8, and a carbonate buffer at pH 9–11. The values of absorbance were measured by UV–Vis spectrophotometer. The pH was adjusted using 0,1 N HCl and 0,1 N NaOH.
**Determination of optimum temperature:** Synthesis of the CuO NPs was carried out separately from 10 °C to 90 °C, respectively, and the changes in absorbance in the samples was measured by UV–VIS spectrophotometer.

**Determination of optimum concentration of metal ion:** Synthesis of the CuO NPs was performed using a copper(II) chloride solution at 0.5 mM, 1 mM, 3 mM, 5 mM and 7 mM, and the absorbance of the samples was measured by a UV–Vis spectrophotometer (350 nm).

**Antimicrobial Activity of Copper Nanoparticles**  
Antagonistic activity of the synthesized CuO NPs was determined against *Pediococcus acidilactici*. A sterile Potato Dextrose Agar (PDA-Oxoid) nutrient medium was prepared and sterilized overnight. The bacterium was studied in three replications, and the average inhibition zone created by the bio-agent was identified with the help of the values obtained [19-20]. In addition, the minimum inhibitory concentration (MIC), defined as the lowest concentration of material that inhibits the growth of a bacterium, was determined as based on batch cultures containing varying concentrations of CuO NPs.

**RESULTS AND DISCUSSION**

**Synthesis of copper nanoparticles**  
300 μg/mL of purified peroxidase enzyme was added in sample of copper(II) chloride solution CuCl$_2$ (10 mM) and incubated in a closed space for 3 hours. The solution was becoming blue to intense blue, which indicates the presence of copper nanoparticles. Then, water was removed with the help of an evaporator and copper nanoparticles were synthesized and they were dried at 65 °C for 48 hours (Figure 1) [21].

![Figure 1. Reaction of green synthesis of copper nanoparticles (A): Before reaction, (B) After reaction.](image)

**Chemical properties of CuO NPs synthesis**  
Contact time was determined to be 30 min., optimum pH was determined as 8.0, optimum temperature was 25 °C, and metal ion concentration was determined as 1 mM for the purpose of optimization of synthesized CuO nanoparticles using a UV-Vis spectrophotometer at 350 nm.
XRD studies
Copper nanoparticles’ XRD which was produced in its peroxidase enzyme catalyst and its crystallographic analysis are given in Figure 2. Characteristic peaks which belong to XRD spectrum in its at $2\theta= 19.86^\circ$, $24.55^\circ$, $58.71^\circ$, $75.22^\circ$ that can be indexed at (111), (200), (220) facets which agree with the values reported for face centered cubic (fcc) copper nanocrystals (Figure 2).

![X-ray diffraction pattern of the synthesis of copper nanoparticles using peroxidase enzyme from Fig (Ficus Carica).](image)

**Figure 2.** X-ray diffraction pattern of the synthesis of copper nanoparticles using peroxidase enzyme from Fig (Ficus Carica).

Surface characterization of CuO NPs
Chemical and mineralogical compositions of synthesized green copper NPs were determined by scanning electron microscopy (SEM), which was used to examine the surface of the adsorbent. Images of CuO NPs were magnified 10000 times by Zeiss, Active area 10 mm$^2$, (Figure 3). It was observed from figure that most of the CuO NPs were spherical in shape. Figure 3 showed a well dispersed CuO NPs had identified in the sizes range 50-120 nm. In CuO nano particles, which were synthesized by using many plant extracts were identified as mostly in spherical from and between 5-200 nm in sizes [21].
Figure 3. SEM image of the synthesized copper nanoparticles.

**Antioxidant activity of CuO NPs**

**The Fe$^{3+}$- Fe$^{2+}$ reducing (FRAP) activity:** In Fe$^{3+}$- Fe$^{2+}$ reducing method, antioxidants give electron and show activity. It is found that the potential of synthesized copper NPs to reduce ferric ions (Fe$^{3+}$) to ferrous ions (Fe$^{2+}$) directly increase in directly proportional with concentration. The reduction was monitored by measuring the change of absorbance at 593 nm. (Figure 4).

![Graph showing FRAP activity](image)

**Figure 4.** The Fe$^{3+}$- Fe$^{2+}$ reducing activity of different concentrations (10-30 µg/mL) of water extracts, alcohol extracts, α−tocopherol, and BHA.

**The Cu$^{2+}$- Cu$^{+}$ reducing activity**

Another method to determine reducing capacity is CUPRAC method. Cupric ions (Cu$^{2+}$) reducing capacity of copper NPs is determined with spectrophotometric method having different concentrations (10-30 µg/mL) of copper NPs. Reducing capacity of (Cu$^{2+}$) by synthesized copper NPs are compared with BHT and α-tocopherol as standard antioxidants and were shown in Figure
5. In 50 μg/mL concentration of copper NPs, capacity of reducing highest cupric ions to (Cu$^{2+}$) cuprous ions (Cu$^{+}$) is found to be BHA $>$ α-tocopherol $>$ copper NP when compared with standards.

![Graph](image)

**Figure 5.** The Cu$^{2+}$- Cu$^{+}$ reducing activity of Copper NPs, BHA and α-tocopherol at different concentrations (10-50 μg/mL).

Superoxide anion radical scavenging activity

One of free radicals, superoxide radical is the easiest and most formed oxygen radical by environmental factors and enzymatic and non-enzymatic reactions in the organism. Superoxide anion radicals cause lipid peroxidation. These radicals cause lipid peroxidation and connected deterioration in membrane structure [22]. Besides, superoxide anion radicals can reduce Fe$^{3+}$ ions to Fe$^{2+}$. Fe$^{2+}$ ions react with Fenton and use hydrogen peroxide to create OH radicals which are very reactive and cause much structural deterioration. Therefore it is necessary to scavenge superoxide anion radicals in the medium. Standard antioxidant materials’ activity to scavenge superoxide anion radical in 50 μg/mL concentration is found to be respectively BHA $>$ α-Tocopherol $>$ Copper NPs. These values respectively are 72.3 ± 0.5 $>$ 56.4 ± 1.1 $>$ 48.3± 1.03. When findings are compared with standards, it is observed that copper NPs scavenged superoxide radicals effectively (Figure 6).
Figure 6. The activity of superoxide anion scavenging of Copper NPs, BHA and α-tocopherol at different concentrations (50 µg/mL)

The ABTS$^{•+}$ scavenging activity
ABTS$^{•+}$ radical is a colored compound which gives absorbance at 734 nm. ABTS$^{•+}$ radical reacts with antioxidant materials and transfers electron and transforms to non-radical ABTS material. Related material is shown below:

\[
\text{ABTS}^{•+} + A \rightarrow \text{ABTS} + A^{•+}
\]

The study used spectrophotometric measurement and followed reducing of absorbance value at 734 nm and calculate ABTS$^{•+}$ radical scavenging activity. ABTS$^{•+}$ scavenging activity is frequently used in radical scavenging activities of liquid mixtures, drinks, extracts and pure materials [23]. Firstly, standard plot is formed to determine scavenging activity of copper NPs produced with green synthesis and standard antioxidant compounds like BHA and α-tocopherol used in study, and ABTS$^{•+}$ scavenging activities are calculated in all samples using standard plot. ABTS$^{•+}$ scavenging activity of copper NPs is compared with standard BHA and α-tocopherol. According to findings, at 50 µg/mL concentration, BHA achieved 73.7%, α-tocopherol achieved 79.7% and copper NPs achieved 85.4% ABTS$^{•+}$ radical scavenging activity (Figure 7). It was found in this study that copper NPs scavenge ABTS$^{•+}$ radical better than standard antioxidants.
Antimicrobial activity of CuO NPs

In the studies, antibacterial features of CuO NPs which were formed with green synthesis method are determined using spread and culture method which is a simple and fast method on nutrient agar. For this purpose, *Pediococcus acidilactici* bacteria is applied on agar surface with spread culture method and it is seen that bacteria growth is avoided around disk and diameter of created inhibition is 16.5 mm. The minimum inhibitory concentration (MIC) value was found Cu NPs (10 µg/mL) using batch method for *Pediococcus acidilactici*. It is understood that copper NPs connect to cell wall of bacteria, damages its structure and cancels its development [24].

CONCLUSION

As a result of our examinations, we understood that peroxidase enzyme can be partly purified from fig leaves (*Ficus carica*) to be used in reaction medium for green synthesis. CuO NPs were characterized using analysis of UV-Vis spectrophotometry, SEM, and XRD. The analysis revealed that the synthesis of CuO NPs and was achieved quantity, direction, and morphology characterization. Manufactured CuO NPs are considered to have been an extensive range of applications in the nanotechnology, the catalyst, pharmaceutical and medical industries. In addition, the experiments showed that CuO NPs have both antimicrobial and antioxidant features. CuO NPs which are acquired with green nano synthesis can be produced more economically and in adequate conditions nontoxic effects on the environment. Besides, examination of electrical and chemical features of acquired CuO NPs can indicate their usage in different areas.

REFERENCES


Türkçe Öz ve Anahtar Kelimeler

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Öz: Bu çalışmanın amacı, kısmen incir yapraklarından (*Ficus carica*) saflaştırılmış peroksidaz enzimlerini kullanarak yeşil bir sentez yöntemi üzerinden bakır oksit nanoparçacıklar (CuO NPler) elde etmektir. CuO NPler, gerçekleştirilen deneylerde yeşil sentez yöntemi ile başarı ile sentezlenmiştir. Elde edilen CuO NPlerin morötesi-görünü (UV-Vis) spektrofotometri ile karakteristik özellikleri, ayrıca taramalı elektron mikroskopisi (SEM) ve X-ışını saçılması (XRD) yapılmıştır. Yeşil sentez için en uygun aktifleşme sıcaklığı 30 dakikada gözlenmiştir, pH 8’dir, sıcaklık 25 °C'dir ve 1 mM CuCl₂ kullanılmıştır. Peroksidaz enzimlerinin yeşil sentezde kullanılması ve SEM ve XRD ölçümlerinin sonuçlarına göre CuO NPlerin boyutlarının 50-120 nm arasında olduğu bulunmuştur. Bunun dışında, bu nanoparçacıkların antioksidan ve antibakteriyel özellikleri ölçülmüş ve CuO NPlerinin antioksidan ve antimikrobiyal aktiviteye sahip olduğu bulunmuştur.

Anahtar kelimeler: Bakır nanoparçacıklar; yeşil sentez; peroksidaz.
