Use of lichens as natural insecticide

Bugrahan EMSEN1*, Ali ASLAN2
1Karmanoğu Mehmetey University, Kamil Özdağ Science Faculty, Department of Biology, Karaman, Turkey
2Yucuncu Yil University, Faculty of Pharmacy, Department of Pharmacology, Van, Turkey
*bugrahanemsen@gmail.com

Abstract: Agriculture has become one of the greatest sources of employment for mankind from the past to the present. The products obtained in this area provide a significant contribution to the national economies. However, the number of organisms causing the decline in crop yield is quite high. The preferred applications for combating harmful organisms are the use of chemical insecticides. However, the high level of side effects of these insecticides led researchers to alternative study areas. Insecticide production through natural products that is under the title of biological struggle, is within popular areas. Lichens are also materials for natural insecticide sources due to their unique constituents in the constructions. For many years, the toxic effects of lichens utilized in many fields on different insect species have been tested through their extracts and pure components. At this point, it is noteworthy that while high yield is obtained from lichens, the studies are predominantly carried out on the extracts. In the light of all these studies, it will be contributed to biological insecticide production stage by increasing of the studies performed on determination of the active components in lichens showing insecticidal activity on different species and on mechanisms of action in insects.

Key words: Extract, Insecticide, Lichen, Metabolite


Anahtar Kelimeler: Ekstrakt, Insektisit, Lichen, Metabolit

1. Introduction

The goal in all past studies is to develop insecticides that can only be effective on the harmful target organism, taking care not to disturb the natural balance without affecting the environment and human health. In this direction, the researchers have been working in the last years and serious studies have started to be done on herbal insecticides (Pant et al., 2016). Natural herbicide insecticides constitute 1% of the world insecticide market. As more work is done on organic agriculture than in the past, annual sales increase by about 10-15% per year (Isman, 1997).

Many researchers have proved that plants are potential sources of insecticides. It was reported that many plant species affect various forms of insects which are harmful to agriculture. Despite the fact that so many of the plants are known to have the insecticidal effect, few have been used in practice. Isman (1997) cites the reasons for this, as natural resources are limited, standardization and licensing difficulties. Natural insecticides have some advantages and disadvantages compared to synthetic insecticides. Natural insecticides break down very quickly in sunlight, humidity and windy conditions, so they can be used just before harvesting. However, many of these insecticides stop feeding their insects very quickly, although they do not kill them immediately. Therefore, it sometimes takes a few days for the death of the insects, but it has a rapid effect in terms of prevention of damage (Oberemok et al., 2015).

If we exclude some highly toxic plant-based insecticides, it can be said that most of the plant-derived natural insecticides are not toxic to mammals and the environment. Rapid disintegration and stomach poisoning make natural insecticides more selective against plant-feeding harmful insects (Walia et al., 2017). Natural insecticides are not usually phytotoxic. They also have no negative effects on their germination, growth and product quality. However, nicotine may have a negative effect on some ornamental plants. When they are used in their natural form, the risks of creating durability are practically absent. They can be obtained cheaper and easier than synthetic insecticides in those areas, as they grow mostly in areas with less developed and lower labor costs (Siervo et al., 2013).

The compounds that plants cause to act on the insects are secondary metabolites that secrete at the time of stress (Rattan, 2010). Herbal substances used against insects are compounds which are obtained by various methods from plants and show insecticidal properties. These can be in various forms, such as unprocessed plant materials, plant extracts, and pure compounds isolated from plants. While some of the natural insecticides are used directly as lethal,.
others have repellent and nourishment-inhibiting properties (Murugesan et al., 2016).

The action mechanisms of insecticides are generally on the nervous and digestive system of the organism. Insecticides of plant origin are said to have entered the body of insects through contact and poisoned the nervous-muscle system via secondary metabolites (Denecke et al., 2015). In addition, it has also been found that herbal insecticides affect physiology of the insect by acting in different forms on various receptor sites (Buckingham et al., 2017).

It is known that plant-based insecticides have effect a number of harmful species. Yildirim et al. (2005) have investigated the effect of three essential oil extracts on Tribolium confusum (du Val) which damages products such as flour and flour produced pasta, biscuits, bran, soup material, starch and some oil seeds and Stiophilus granarius (L.) damaging in stored seed known as wheat weevil. As a result of the experiment, mortality rate of 67% for T. confusum and 43% for S. granarius appeared. In another study, fumigant toxicity effect of essential oils from nine plant species on adults of S. granarius was investigated (Kordali et al., 2012).

Yildirim et al. (2009) have evaluated effectiveness of plants grown from pesticide seeds against cabbage aphid (Brevicoryne brassicae L.), cabbage leaf fleas (Phyllotreta atra F., Phyllotreta nigripes F.), cabbage moth (Plutella xylostella L.) and large cabbage butterfly (Pieris brassicae L.). As a result, it was detected that the damage of leaf fleas in cabbage plants was considerably reduced and after six weeks, the loss was reduced by 90%. Many studies that demonstrate that some medical plants have the same effect as some chemical insecticides are also included in the literature (Tewary et al., 2005; Kordali et al., 2013; Yildirim et al., 2013). In recent years, insecticidal activity studies carried out through compounds derived from in vitro propagated plants have also gained popularity (Emsen et al., 2016; Dogan et al., 2017).

Since plant origin ones among the insecticides used in the struggle with harmful organisms act differently on the target organism without harming the environment, their usage areas have expanded.

2. General Characteristics of Lichens

Among the many species of organisms used for the purpose of obtaining insecticides, lichens are the most remarkable species. Lichens are very interesting livings with their forms of formation, their living habits, their diversity of uses, and their components that they have in their structure. According to Nash (2008), lichens are symbiotic organisms formed with a fungus partner called a mycobiont and by the photosynthetic partner which can be one or more algae or cyanobacteria and is known as a photobion. Lichens show a completely separate structure from the algae and fungi that form themselves in terms of shape and life. The photosynthetic organism (photobiont) involved in the structure of a colorless fungal hyphae is usually green algae or a cyanobacterium; but they are also known to be composed of some yellow-green algae and brown algae (Bačkor and Fahselt, 2008). Some of the genus belonging to Cyanophyta and Chlorophyta and some algae from Xanthophyta and Phaeophyta are observed. Mushrooms are usually Ascomycetes and rarely belonging to Basidiomycetes (Grube and Spribille, 2012; Millot et al., 2016).

In the periods when the lichens first began to be examined, although the fungus in the lichen symbiosis was thought to have benefited from algae such as parasites due to lack of chlorophyll, but later this explanation lost its significance. It has been understood that the fungus has been supplied with water and water-soluble minerals from the environment in common life with the help of hyphae (Brodo et al., 2001; Nash, 2008). Fungi that can not produce their own carbohydrates meet the needs for glucose from photosynthesizing algae and cyanobacteria thanks to the chlorophyll pigment contained. Starch is also used as a storage material in lichens (Rikkinen, 2015; Suzuki et al., 2016).

For years, many scientists have gone to the way of classifying lichens by adding their own interpretation. One of these classifications is based on the environment on which lichens live. According to this, lichens are divided as living on limestone rocks, siliceous soils, tree trunks and branch bark, leaves, overturned tree logs, and in water. Another classification has been made according to the type of fungus found in the lichen species. In this case, lichens are divided into three classes as Phycocichens, Ascolichens and Basidiolichens (Brodo et al., 2001). Nowadays, the most important classification accepted by many scientists is to group lichens according to thallus structures. The thallus structures found in lichens show the character of this lichen. Because, in many cases, a great deal of a lichen comes from the thallus, unlike the reproductive and reproduction processes. According to thallus structure, lichens are divided into three groups, namely crustose, foliose and fruticose (Sipman, 2002; Armstrong and Bradwell, 2010).

Among the lichens, the species that are short-lived are rare. Although there are environmental issues that reduce the lifespan of lichens like other life forms, they can survive for more than 1000 years under favorable conditions such as fresh air, adequate humidity and light (Armstrong, 2004). Lichen species are very susceptible to polluted air, so the lichen flora is very poor in industrial areas and near large cities. On the other hand, the lichens cover the rocks, tree trunks and branches in various colors and shapes in clean air areas. For this reason, they are indicator organisms for the cleanliness of a zone's air (Szczepaniak and Biziuk, 2003; Rikkinen, 2015).

Lichens are distributed almost everywhere in the world. They are able to develop even in the desert where there is enough humidity, Arctic and Antarctic regions, the freezing temperatures of high mountains, on the stones that other plants can not live on, insufficient soil, dry tree bark and tiles (Kranner et al., 2008). The feature that allows lichens to survive on extreme conditions is hidden in the acids in their structure. Lichen acids, the secondary metabolites of lichens, give the lichens the ability to become a preliminary organism in the primary succession in the nature. The sandy, rocky, clayey, marsh and gravel environments are primarily occupied by lichens. These are called leading populations. At this point, lichens increase the soil quality of the environment (Xu et al., 2005; Zambare and Christopher, 2012; Nguyen et al., 2013).
3. Different Usage Areas of Lichens

Due to the lichen acids produced by the fungi in the symbiotic life of lichens, since the past centuries it was benefitted from the lichens in many areas. For example; in the past, during scarcity in Scandinavia, Cetraria islandica (L.) Ach. was milled and added to wheat flour or potatoes to increase insufficient supplies. In the 1880s in Sweden, Cladina rangiferina (L.) Nyl. was used as a sugar source (Brodo et al., 2001). C. islandica and Lobaria pulmonaria (L.) Hoffm. have been used for skin tanning because of their shrunken properties. L. pulmonaria has also been used in the fermentation of beer. It has been found that the beer made with this lichen is exactly the same as the one made with hops (Tutel, 1986).

Most of the Roccella species from lichens were used in the dyeing of wool and clothes until the mid-19th century. Two different color substances were identified among Roccella species. There is erythrin acid in Roccella fuciformis (L.) DC., Roccella fucoides (Dicks.) Vain., Roccella perennis Krempl., Lecanora montagnei (Fr.) Schair. and lecanorin acid was determined in Roccella tinctoria DC., Roccella sinsensis Nyl. and Cladonia portentosa ((Dufour) Coem.) (Aslan, 1995; Culberson and Culberson, 2001). Evernia pruinosa (L.) Ach., Pseudevernia furfuracea (L.) Zopf., L. Pulmonaria, Anaptychia ciliaris (L.) Körb. and some Usnea sp., Physcia sp. were used in the field of cosmetics. These lichens are used in perfume as a mixture, not alone. The lichens that increase the persistence of smell give a pleasant fragrance to the combination of perfume (Tutel, 1986).

The lichens were also used in the medical field. The lichen species in this area are generally used with meaning according to their morphological appearance. L. pulmonaria was used as a softening cream in the treatment of pulmonary disease for pulmonary resemblance. Usnea Florida (L.) Weber ex F.H. Wigg. and many other Usnea species, which are filamentous and resembling hair, were used to prevent hair loss and to make hair look louder and more vibrant. Xanthoria parietina (L.) Beltr. which has a yellow-orange color and Peltigera aphthosa (L.) Willd., which is similar to the small warts in the thallus were used in the treatment of jaundice and canker diseases, respectively (Brodo et al., 2001).

Lichens are used in the form of laxatives, expectorants, tonic, internal or external pastes, decoction (boiling) or infusion. In various parts of the world, researches are being conducted for new pharmaceutical uses of lichens (Shukla et al., 2010). It has been found that some lichens have antitumor activity through components such as polysaccharide, glucan and glycoprotein. Antimicrobial activities against different bacterial species were identified in lichens where many antibiotic substances were detected (Shrestha and St. Clair, 2013; Shrestha et al., 2015).

4. Insecticidal Efficacies of Lichens

It is also necessary to add the insecticide activities to the above-mentioned and many other unused usage areas of lichens. It was reported that Letharia vulpina (L.) Hue ve Vulpicia pinastri (Scop.) J.-E. Mattsson species are toxic and these lichens have been used in some European and Scandinavian countries to kill wolves and foxes that have damaged to animals in winter . When these species are eaten by animals, they cause death by stopping breathing, particularly by influencing the respiratory system. Brodo et al. (2001) reported that a lichenologist who collected L. vulpina from these toxic species in large quantities severely undergone respiratory tract irritation since he was constantly in contact with this species and this person had a nosebleed. This event suggests that these lichen species may also be poisonous to humans. Some researchers, considering these properties of lichens, have gone to investigate the insecticidal effect of lichens.

Cetin et al. (2008) investigated the insecticidal effects of (-)-usnic acid and (+)-usnic acid secondary metabolites obtained from Cladonia foliacea (Huds.) Wield. and Ramalina farinacea (L.) Ach. against Culex pipiens L. (mosquito) larvae in laboratory conditions and get 100% results in some biological periods of these organisms. 1440 samples from 50 lichen species with different chemical properties were analyzed to determine whether these characteristics of lichens-fed insects originated from lichen species, particularly lichen chemistry by Nimis and Skert (2006). Multivariate analyzes revealed a special negative correlation between the presence of grass-fed animals and some lichen substances.

Silva et al. (2009) examined potential insecticide effects of lectin isolated from Cladonia verticillaris (Radde) Fries on Nasutitermes corniger Motschulsky termite. As a result of their work, it was stated that C. verticillaris preparations might be able to control termites (or other insects) that are economically related to agriculture and the wood industry. In another study, the effects of methanol, chloroform and water extracts of L. pulmonaria on the life span of Drosophila melanogaster Meigen (vinegar fly) were investigated (Uysal et al., 2009). They found that methanol extract was more effective than chloroform and water extracts and water extract was relatively weaker than methanol and chloroform extracts. Emmerich et al. (1993) investigated the antifeedant and lethal effects of four of the lichen metabolites, (-)- and (+)-usnic acid, vulpinic acid, and stictic acid on Spodoptera littoralis Böisduval larvae. They noted that both types of usnic acid as well as vulpinic acid showed a lethal effect at high levels and delayed growth. It was also detected that stictic acid did not show any effect.

In various treatments, it was reported that larval killer effect of usnic acid and vulpinic acid was tested on Bemisia tabaci Gennadius (white fly) and vulpinic acid was more effective than (-)-usnic acid. In addition, it was determined that hexane, ethyl acetate, methanol and water extracts of Roccella montagnei Bél. had a lethal effect on Helicoverpa armigera (Hubner) Among (green worm) (Balaji et al., 2007). Another lichen-insecticide related study was performed on Usnea sp., Heteroderma diademata (Taylor) D.D. Awasthi, Roccella montagnei Bél. and Leproloma sipmanianum Kümmel & Leuckert, from lichen species. In the related work, high insecticidal effect of ambewelamide A compound isolated from aforementioned lichens was determined on the second period larvae of Aedes aegypti (L.). In another study performed with the same larvae, it was pronounced that cabraleadiol monoacetate and 3,6-dimethyl-2-hydroxy-4-methoxybenzoic acid from lichen acids had insecticidal activity (Tufan Çetin and Sümül, 2008).
In recent years, efforts to create biological insecticides against storage pests through different lichen extracts and components have gained popularity. Emsen et al. (2012b) observed toxicity of Cladonia foliciaea (Huds.) Willd. and Flavoparmelia caperata (L.) Hale on grain weevil, Sitophilus granarius (L.) and reported 91 and 83% mortality rates for both species, respectively. Similarly, experiments carried out with L. vulpina, Lecanora muralis (Schreb.) Rabenh. and Peltigera rufescens (Weiss) Humb revealed other lichen species that was effective on S. granarius (Emsen et al., 2015). In that study where concentration-dependent insecticidal activity was observed, median lethal concentration (LC₅₀) values of L. vulpina, L. muralis and P. rufescens were 0.51, 0.67 and 0.33, respectively. Insecticidal activities caused by extracts of L. vulpina, L. muralis and P. rufescens were 100, 86 and 100%, respectively.

There are also studies in which insecticidal effects of active ingredients isolated from lichens against grain weevil are tested. Yildirim et al. (2012a) obtained diffractaic and usnic acid from Usnea longissima Ach. and measured insecticidal effect of these secondary metabolites. Although they found that both compounds had high activity, it was revealed that usnic acid was more effective than diffractaic acid. Another insect that causes loss in warehouse products is Sitophilus zeamais Motschulsky. Lichens were also used to combat this pest through natural insecticide. Diffractaic acid and usnic acid from pure lichen compounds and L. vulpina, P. rufescens extracts were tested against S. zeamais and positive results achieved (Yildirim et al., 2012b). In previously mentioned study, the highest mortalities belonged to L. vulpina extract and diffractaic acid with 96.97%. L. vulpina, L. muralis and P. rufescens have taken place among lichens used for solution to potato beetle (Leptinotarsa decemlineata Say) that damages an important agricultural product, potato. The insecticidal activities of L. vulpina and P. rufescens after 120 hours was remarkable (Emsen et al., 2013). In the same way, when measured the toxicity efficacies of diffractaic and usnic acids isolated from U. longissima against 4th instar larvae and adults of L. decemlineata, it was determined that the activity levels of mentioned compounds were significant after 96 hours (Emsen et al., 2012a).

The lichens that are used in different areas due to their unique compounds also have a place in the field of biological insecticide. However, total extracts are noteworthy in the studies carried out so far for natural insecticide production field. In the future, increasing of the studies performed on detection of the active components that play insecticidal role in the lichens and on insect death mechanisms will contribute much more to the field of biological insecticide production.

Conflicts of interest
There is no conflict of interest in any form between the authors.

References


